

EFFECTS OF WATER DEFICIT AND SALT STRESS ON SOME PHOTOSYNTHESIS PARAMETERS IN WHEAT AND *AEGILOPS COMOSA* LINES

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Abstract: Photosynthetic responses of *Aegilops comosa* genotypes were compared to those of wheat Mv9kr1 and Chinese spring in order to verify whether *Ae. comosa* TA2760 and MvGB1039 genotypes are potentially suitable gene sources for improving the drought and salt tolerance of bread wheat. Although there are some differences between the non-stressed plants and the measure of the decrease of the net-photosynthesis (P_N), it was strongly inhibited by water deficit. Salt stress had similar effect on P_N but at the highest (300 mM) NaCl concentration P_N of the genotypes showed some activity. Severe drought induced a strong decrease in the effective quantum yield of PS II (ϕ_{PSII}) in the genotypes, while it was moderate in the case of salt treatment. Moreover, ϕ_{PSII} was unaffected by the increase of NaCl concentration in wheat lines. Parallel with the decrease in ϕ_{PSII} , the photoprotective mechanisms were enhanced in the wheat and wild wheat genotypes during water deficit. These results suggest that the *Ae. comosa* genotypes seem to respond to these stress factors with similar photosynthetic activity to the wheat lines. Thus, based on the above-mentioned facts, the examined *Ae. comosa* lines are not particularly good candidates for improving drought tolerance of wheat.

Keywords: wheat, *Aegilops comosa*, photosynthesis, water deficit, salt stress

INTRODUCTION

Abiotic environmental factors due to the forecasted effects of global climate change can considerably endanger the productivity of cultivated plants. An important problem is the rhapsodic rainfall

pattern (Trenberth *et al.* 2007), which often results agricultural damages by drought and/or salt stress. Although drought is the most common abiotic stress factor (Araus *et al.* 2002) but also the agricultural lands are often affected by salt stress (Munns 2005). In field conditions, a positive relationship of photosynthetic capability and crop production has been well documented. At the same time, photosynthesis is particularly sensitive to drought and salinity (Ashraf and Harris 2013, Dulai *et al.* 2014, Szopkó *et al.* 2017a, Szopkó and Dulai 2018).

Under stress conditions the limitation of photosynthetic capacity takes place in two stages: (i) limitation associated with decreased stomatal conductance, known as stomatal limitation (L_s , Centritto *et al.* 2003, Dulai *et al.* 2014); (ii) limitation due to nonstomatal processes (L_{ns}) mainly at severe drought and higher salt concentrations or longer salt stress (James *et al.* 2002, Centritto *et al.* 2003, Munns *et al.* 2006, Szopkó *et al.* 2017b). Under moderate water deficit or in the first stage of salt stress, which also has osmotic effect, the reduction in net photosynthesis (P_N) mainly due to the stomatal closure (Chaves 1991; Medrano *et al.* 2002). In this case the CO_2 diffusion into the leaves is also restricted resulting in a decrease in the intercellular CO_2 concentration (C_i) (Cornic 2000) and CO_2 carboxylation (Flexas *et al.* 2004). At the same time, the reduced stomatal conductance (g_s) may contribute to maintaining water content through a decreased transpiration rate, which could be favourable for minimizing Na^+ transport towards the shoots (Tester and Davenport 2003). Thus, closed stomata have both positive and negative effects on photosynthesis (Szopkó *et al.* 2017b).

When the stress turns severe, photosynthesis is also limited by factors other than stomatal closure. The drought induced non-stomatal limitation of P_N may be caused by the restricted mesophyll conductance (Loreto *et al.* 2003) or by metabolic factors (Medrano *et al.* 1997, Centritto *et al.* 2003, Chaves *et al.* 2003). Similar to water deficit, salt stress also has many consequences for non-stoma-dependent processes as well. Salt-induced non-stomatal inhibition (L_{ns}) can be observed when the CO_2 assimilation is disturbed by the presence of toxic ions. This limitation may be associated with limited Rubisco activity a reduced amount of Rubisco protein or weak efficiency of PSII in the second stage of salt stress (Muranaka *et al.* 2002, Kalaji *et al.* 2011), when a high

concentration of toxic Na⁺ and Cl⁻ ions evolves in the leaves (Munns and Tester 2008).

During drought and salt stress photosynthesis is often hindered by the secondary effect of disturbed water and ion homeostasis. The increasingly severe limitation of photosynthesis leads to the plant absorbing more light energy than that can be used by CO₂ fixation (Smirnov 1993). Although the excess light can be partially dissipated as heat, it has the potential to cause the over-reduction of the linear electron transport chain, leading to oxidative damage (Smirnov 1993, Flexas *et al.* 2004). Under these circumstances the down-regulation of photosynthesis by non-radiative energy dissipation (Demmig-Adams *et al.* 1996) and/or photorespiration represent an efficient defence mechanism in C₃ plants. Thus, the facility to maintain promising photosynthesis and consequently achieve adequate growth and production are based on these intensive protecting/regulating mechanisms.

Interspecific hybridization of wheat with wild relatives is an appropriate breeding strategy to improve the stress tolerance (Colmer *et al.* 2006, Schneider *et al.* 2008, Pradhan *et al.* 2012). *Aegilops* species are widely used as genetic resources in the breeding of bread wheat. These plants are native in the Mediterranean coastal areas characterised by hot, dry vegetation periods often with high salinity (Molnár *et al.* 2004, Dulai *et al.* 2006). Relating to this, *Aegilops* species might adapt to the unfavourable environmental conditions thus their ability to tolerate some abiotic stresses has already been partly described (Zaharieva *et al.* 2001, Molnár *et al.* 2004, Dulai *et al.* 2006).

However, the drought and salt tolerance of some *Aegilops comosa* genotypes is unclear. The aim of the present study was to clarify the drought and salt tolerance of the *Ae. comosa* TA2760 and MvGB1039 lines. For this purpose these plants were exposed to drought and salt stress and the photosynthetic responses of these genotypes were compared to those of wheat Mv9kr1 and Chinese spring. It is revealed that whether *Ae. comosa* TA2760 and MvGB1039 genotypes are potentially suitable gene sources for improving the drought and salt tolerance of bread wheat.

MATERIALS AND METHODS

Plant materials

In our experiments *Ae. comosa* TA2760, MvGB1039, Mv9kr1 and Chinese spring wheat genotypes were investigated. The seeds of lines were provided by Márta Molnár-Láng and István Molnár, Agricultural Institute of the Hungarian Academy of Science (Martonvásár, Hungary). The seeds were germinated in laboratory conditions on filter paper moistened with distilled water than they were grown in half-strength modified Hoagland nutrient solution (Nagy and Galiba 1995) in 1500 ml pots or were planted in soil (5 seeds/pot). Plants grow at 25/20°C in a growth chamber with a photosynthetic photon flux density of 200 $\mu\text{E m}^{-2} \text{s}^{-1}$ and 14/10 hours of light/dark illumination. Salt stress was induced by applying 150 and 300 mM concentration of NaCl (Sigma, St. Louis, USA) in seven-day cycles. After reaching the highest salt concentration, the salt was eliminated from the medium. Measurements were made before the treatment (5-week old plant), after each seven-day treatment and after two and seven days of regeneration without NaCl. The watering of the *Aegilops* lines and wheat lines grown in soil was abolished after the age of 5 weeks. In the case of water-deficient plants, the measurements were performed on the 4th, 7th and 10th day of water shortage. The regeneration ability was investigated by the total humidification of the soil.

Gas exchange measurements

The CO_2 assimilation of intact leaves was measured with an infrared gas analyser (GFS-3000FL, Walz, Effeltrich, Germany). The net assimilation rate (P_N) was calculated in the light-saturated state of photosynthesis ($1000 \mu\text{mol m}^{-2} \text{s}^{-1}$) using the equations reported by von Caemmerer and Farquhar (1981). The gas exchange chamber parameters were 25°C, 20% relative humidity. The CO_2 concentration of the reference air was 360 ppm.

Chlorophyll fluorescence measurements

The *in vivo* chlorophyll *a* fluorescence was measured in dark-adapted intact leaves using a dual channel P700 and chlorophyll fluorescence measuring system (Dual PAM-100, Walz, Effeltrich, Germany). The initial level of fluorescence (F_0) was excited by a

weak 460-nm light beam after 15 min dark adaptation. The maximal fluorescence level of the dark- (F_m) and light- (F_m') adapted leaves were determined by applying saturating flashes ($15000 \mu\text{mol m}^{-2} \text{s}^{-1}$ PAR) with 0.8 s duration. Photosynthesis was induced by continuous illumination of the leaf at $1000 \mu\text{mol m}^{-2} \text{s}^{-1}$ (650 nm, actinic light) for 10 min. The fluorescence parameters were calculated as described by van Kooten and Snel (1990) on the basis of the following equations: maximal quantum yield of PSII, $F_v/F_m = (F_m - F_0)/F_m$; effective quantum yield of PSII, $\Phi_{\text{PSII}} = (F_m' - F_s)/F_m'$; non-photochemical quenching, $\text{NPQ} = (F_m - F_m')/F_m'$.

Statistical analysis

Five measurements were performed on each genotypes and treatment for chlorophyll fluorescence and for CO_2 gas exchange analyses. The results are presented as the means \pm standard deviations (SD) of five independent experiments. Differences between treatments or genotypes within each treatment were determined by means of Tukey's post hoc test ($p \leq 0.05$) using the SPSS 16.0 software.

RESULTS

Water deprivation, similar to increase of salt concentration, resulted in a gradual decrease in stomatal conductance. In the water stressed lines almost total closure of the stomata was observed at the 10-day water stress while it was not fully complete at the highest salt concentration (data not shown). Although there are some differences between the non-stressed plants and the measure of the decrease of the CO_2 assimilation rate, P_N was strongly inhibited by water deficit at the seventh day of treating (*Figure 1A*). When the drought stress was more severe (at the tenth day of the treatment) P_N was almost fully inhibited in all of the examined genotypes. Salt stress had similar effect on P_N but at the highest (300 mM) NaCl concentration, P_N of the genotypes showed some residual activity. However, the measure of this inhibition was more or less proportional to that of the water-deficient plants. During the regeneration period P_N was restored swiftly for the drought treated plants: the recovery was complete on the second day of the relaxation period. In the case of salt stress the

restoration was slower but also complete by the 7th day of the regeneration period.

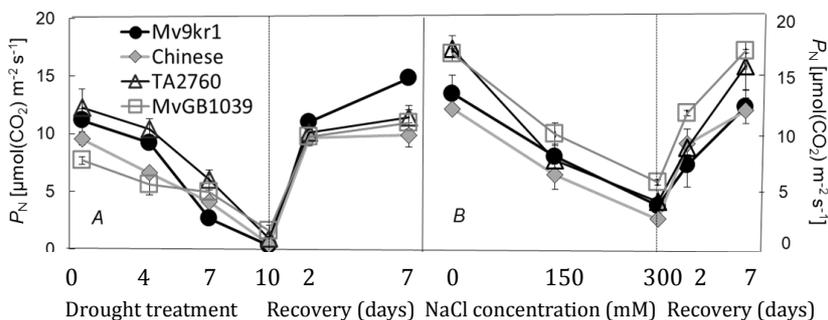


Figure 1. Effects of water deficit, NaCl concentration and recovery on net photosynthesis (P_N) in wheat (Mv9kr1, Chinese spring) cultivars and *Aegilops comosa* (TA2760, MvGB1039) genotypes.

Chlorophyll fluorescence measurements provide a fast and adequate method for analysing the functioning of the photosynthetic apparatus. Although there was a slight decrease in the optimal quantum yield of PS II (F_v/F_m) by the second day of the regeneration period in the case of the salt treatment, both water and salt stresses did not result a significant decrease in this parameter in any plants (Figure 2C, F). In untreated plants and under mild (four day drought treatment and 150 mM NaCl) stress conditions, ϕ_{PSII} was similar in the genotypes (Figure 2A, D). The severe drought induced a strong decrease in the effective quantum yield of PS II (ϕ_{PSII}) in the genotypes, while it was moderate in the case of salt treatment. Moreover, ϕ_{PSII} was unaffected by the increase of NaCl concentration in wheat Mv9kr1. Parallel with the decrease in ϕ_{PSII} , the non-photochemical quenching (NPQ), reflecting the regulated heat dissipation, increased especially during the drought treatment, and slowly recovered to the original level after the stress treatments (Figure 2B). By contrast, NPQ showed a slight increase in Mv9kr1 and Chinese spring wheat cultivars during salt treatment. At the same time, this parameter rose sharply in *Ae. comosa* TA2760 and MvGB1039 and did not recovered fully during the seven days relaxation period (Figure 2E). Thus, the 10-day water deprivation induced NPQ in all lines with very similar extent, but the increase of photo-protective

mechanisms were detected only in *Ae. comosa* genotypes when salt stress was developed.

DISCUSSION

Under water deficit and/or salt stress, the stomata closure is a well-known phenomenon (Molnár *et al.* 2004, Dulai *et al.* 2014, Szopkó *et al.* 2017b). The decrease in stomatal conductance (g_s) affects not only the regulation of water loss through the transpiration, but also inhibits the photosynthetic CO₂ fixation by limiting CO₂ diffusion into the leaves (Chaves 1991, Cornic 2000, Flexas and Medrano 2002). As mentioned above, the water deprivation caused an almost total closure of the stomata at the 10-day water stress (data not shown). In our experiments, the CO₂ assimilation rate modified likewise as g_s : P_N values were substantially decreased not only in wheat but also in wild wheat lines. Although the better tolerance of several *Aegilops* species to drought is well documented (Molnár *et al.* 2004, Dulai *et al.* 2006), these results indicate that drought tolerance of the examined wild wheat lines according to CO₂ assimilation processes is similar to those in wheat genotypes. Thus, based on the overall photosynthetic capacity, the examined *Ae. comosa* lines are not particularly good candidates for improving drought tolerance of wheat.

Similar to drought, photosynthetic processes are also modified during salt stress (Szopkó *et al.* 2017b). Prior to the accumulation of toxic ions, salt treatment also causes osmotic stress, influencing the water status, stomatal conductance and net carbon fixation capacity of plants (Munns 2002, Munns and Tester 2008). P_N was a little bit higher in *Ae. comosa* MvGB1039 than this was in wheat lines at all levels of salt treatment ($P \leq 0.05$). Apart from this, it decreased considerably even at a moderate stress level in all lines.

These results show that the examined lines were not able to maintain their photosynthesis at a promising level during salt stress. Compared to the drought treatment, at 300 mM salt concentration P_N of the genotypes showed a little bit higher activity than the 10-day water deprivation. This is probably due to the less strong closure of stomata.

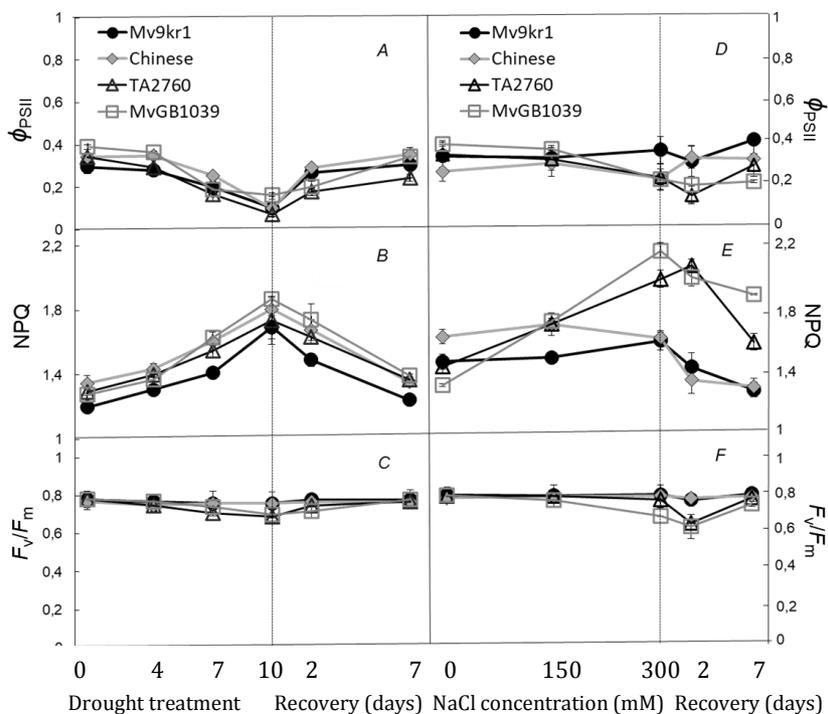


Figure 2. Effects of water deficit, NaCl concentration and recovery on effective quantum yield (A, D), non-photochemical quenching (B, E) and optimal quantum yield of PS II (C, F) in wheat (Mv9kr1, Chinese spring) cultivars and *Aegilops comosa* (TA2760, MvGB1039) genotypes.

When drought or salt stress become severe and CO_2 assimilation is strongly inhibited, the role of non-stomatal factors in the limitation of photosynthesis usually becomes more pronounced (Brugnoli and Lauteri 1991, Qin *et al.* 2010). One of the non-stomatal factors affecting CO_2 fixation during water deficit or salt stress is the inhibition of the photochemical and electron transport processes (Keck and Boyer 1974, Giardi *et al.* 1996, Szopkó *et al.* 2017b). At the same time, the contribution of these processes to the limitation of P_N usually depends on the duration/intensity of the treatment (Kalaji *et al.* 2011). In the present experiments, the optimal quantum yield (F_v/F_m) were practically unaffected by the treatments. Consequently, our results show that the applied water deficit and salt treatment has only a marginal effect on the capacity of primary charge separation. Thus, no PSII damage observed, as

also reported by previous studies (Ben *et al.* 1987, Grieu *et al.* 1995, Dulai *et al.* 2006, 2014, Stiller *et al.* 2008, Szopkó *et al.* 2017b).

The different performance of effective quantum yield of PSII (ϕ_{PSII}), indicated that the electron transport processes were influenced distinctly by drought and salt stress in the given lines. Drought has significant effect on ϕ_{PSII} in all lines: the values of this parameter gradually decreased during the treatment indicating that electron transport processes were partly down-regulated in these genotypes. Parallel with the decrease of ϕ_{PSII} , the photoprotective mechanisms were intensely accelerated in the wheat and wild wheat genotypes during water deficit, as indicated by the higher values of the non-photochemical quenching (NPQ). These processes compete with primary photochemistry for the absorbed excitation energy, leading to a decrease in ϕ_{PSII} (Genty *et al.* 1989) and an increase in non-radiative energy dissipation in the light-harvesting complexes (Horton and Ruban 2005, Chaves *et al.* 2009). In spite of the drought treatment, ϕ_{PSII} was less sensitive to the applied range of salt stress in most of the lines, and even in wheat genotypes, it has not decreased in parallel with the treatment. It seems unlikely that the down-regulation of the PSII-driven electron transport is responsible for the limitation of photosynthesis, because the decrease in ϕ_{PSII} was relatively moderate and the relaxation of photosynthetic CO₂ fixation in wheat genotypes was faster than the recovery of ϕ_{PSII} . It has also been shown by several authors (Apostolova *et al.* 2006, Dulai *et al.* 2014, Szopkó *et al.* 2017b) that PSII is usually more sensitive to stress factors (drought, salt, heat, etc.) than PSI. Moreover, PSI activity may even be enhanced during salt stress (Sudhir *et al.* 2005). Consequently, there is a possibility that electrons may also follow a cyclic route, around the PSI and known as cyclic electron flow (CEF), which generate ΔpH across the thylakoid membranes leading to the formation of ATP but not NADPH, thus preventing the over-reduction of the acceptor side of PSI. Consequently, CEF helps to prevent the subsequent oxidative damage when carbon fixation is limited by water deficit or salt stress (Golding and Johnson 2003, Dulai *et al.* 2014, Szopkó *et al.* 2017b). Based on the higher values of NPQ in all lines under severe drought and in *Aegilops* genotypes under severe salt stress might make it possible that CEF may help to prevent the over-reduction of the electron

transport chain and subsequent oxidative damage (Dulai *et al.* 2014, Szopkó *et al.* 2017b).

In conclusion, the results proved that the examined *Aegilops comosa* lines were not able to maintain their photosynthesis at a promising activity under drought and salt stress. These lines seem to respond to these stress factors with similar photosynthetic activity to the wheat lines. Thus, based on the above-mentioned facts, the examined *Ae. comosa* lines are not particularly good candidates for improving drought and/or salt tolerance of wheat.

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REFERENCES

- APOSTOLOVA, E.L., DOBRIKOVA, A.G., IVANOVA, P.I. PETKANICHIN, I.B. & TANEVA, S.G. (2006). Relationship between the organization of the PSII supercomplex and the functions of the photosynthetic apparatus. *Journal of Photochemistry and Photobiology B* **83**: 114–122.
<https://doi.org/10.1016/j.jphotobiol.2005.12.012>
- ARAUS, J.L., SLAFER, G.A., REYNOLDS, M.P. & ROYO, C. (2002). Plant breeding and water relations in C3 cereals: what should we breed for? *Annals of Botany* **89**: 925–940. <https://doi.org/10.1093/aob/mcf049>
- ASHRAF, M. & HARRIS, P.J.C. (2013). Photosynthesis under stressful environments: An overview. *Photosynthetica* **51**: 163–190.
<http://dx.doi.org/10.1007/s11099-013-0021-6>
- BEN, G.Y., OSMOND, C. & SHARKEY, T. (1987). *Effects of water stress on in vivo photosynthetic biochemistry (maximum photosynthesis quantum yield and 77K fluorescence)*. In: BIGGINS J. (ed.): *Progress in Photosynthesis Research*. Vol. V. Martinus-Nijhoff Publisher, Dordrecht, pp. 157–160.
- BRUGNOLI, E. & LAUTERI, M. (1991). Effects of salinity on stomatal conductance, photosynthetic capacity, and carbon isotope discrimination of salt-tolerant (*Gossypium hirsutum* L.) and salt-sensitive (*Phaseolus vulgaris* L.) C₃ non-halophytes. *Plant Physiology* **95**: 628–635.
<https://doi.org/10.1104/pp.95.2.628>
- CENTRITTO, M., LORETO, F. & CHARTZOULAKIS, K. (2003). The use of low [CO₂] to estimate diffusional and non-diffusional limitations of photosynthetic capacity of salt-stressed olive saplings. *Plant Cell and Environment* **26**: 585–594.
<https://doi.org/10.1046/j.1365-3040.2003.00993.x>
- CHAVES, M.M. (1991). Effects of water deficits on carbon assimilation. *Journal of Experimental Botany* **42**: 1–16. <https://doi.org/10.1093/jxb/42.1.1>
- CHAVES, M.M., FLEXAS, J. & PINHEIRO, C. (2009). Photosynthesis under drought and salt stress: regulation mechanisms from whole plant to cell. *Annals of Botany* **103**: 551–560. <http://dx.doi.org/10.1093/aob/mcn125>

- CHAVES, M.M., MAROCO, J.P. & PEREIRA, J.S. (2003). Understanding plant responses to drought – from genes to whole plant. *Functional Plant Biology* **30**: 239–264. <http://dx.doi.org/10.1071/FP02076>
- COLMER, T.D., FLOWERS, T.J. & MUNNS, R. (2006). Use of wild relatives to improve salt tolerance in wheat. *Journal of Experimental Botany* **57**: 1059–1078. <https://doi.org/10.1093/jxb/erj124>
- CORNIC, G. (2000). Drought stress inhibits photosynthesis by decreased stomatal aperture – not by affecting ATP synthesis. *Trends in Biochemical Sciences* **5**: 187–188. [https://doi.org/10.1016/S1360-1385\(00\)01625-3](https://doi.org/10.1016/S1360-1385(00)01625-3)
- DEMIG-ADAMS, B., ADAMS, III W.W., BARKER, D.H., LOGAN, B.A., BOWLING, D.R. & VERHOEVEN A.S. (1996). Using chlorophyll fluorescence to assess the fraction of absorbed light allocated to thermal dissipation of excess excitation. *Physiologia Plantarum* **98**: 253–264. <https://doi.org/10.1034/j.1399-3054.1996.980206.x>
- DULAI S., MOLNÁR I., SZOPKÓ D., DARKÓ E., VOJTKÓ, A., SASS-GYARMATI, A. & MOLNÁR-LÁNG, M. (2014). Wheat-*Aegilops biuncialis* amphiploids have efficient photosynthesis and biomass production during osmotic stress. *Journal of Plant Physiology* **171**: 509–517. <https://doi.org/10.1016/j.jplph.2013.11.015>
- DULAI, S., MOLNÁR, I., PRÓNAY, J., CSERNÁK, Á., TARNAI, R. & MOLNÁR-LÁNG, M. (2006). Effects of drought on photosynthetic parameters and heat stability of PSII in wheat and in *Aegilops* species originating from dry habitats. *Acta Biologica Szegediensis* **50**: 11–17.
- FLEXAS, J., BOTA, J., LORETO, F., CORNIC, G. & SHARKEY, TD. (2004). Diffusive and metabolic limitations to photosynthesis under drought and salinity in C₃ plants. *Plant Biology* **6**: 269–279. <https://doi.org/10.1055/s-2004-820867>
- FLEXAS, J. & MEDRANO, H. (2002). Drought-inhibition of photosynthesis in C₃ plants: stomatal and non-stomatal limitations revisited. *Annals of Botany* **89**: 183–189. <https://doi.org/10.1093/aob/mcf027>
- GENTY, B., BRIANTAIS, J.M. & BAKER, N.R. (1989). The relationship between the quantum yield of photosynthetic electron transport and quenching of chlorophyll fluorescence. *Biochimica et Biophysica Acta (BBA) – General Subjects* **990**: 87–92. [http://dx.doi.org/10.1016/S0304-4165\(89\)80016-9](http://dx.doi.org/10.1016/S0304-4165(89)80016-9)
- GIARDI, M.T., CONA, A., GIEKEN, B., KUCERA, T., MASOJÍDEK, T. & MATTOO, A.K. (1996). Long-term drought stress induces structural and functional reorganization of photosystem II. *Planta* **199**: 118–125.
- GOLDING, A.J. & JOHNSON, G.N. (2003). Down-regulation of linear and activation of cyclic electron transport during drought. *Planta* **218**: 107–114. <https://doi.org/10.1007/s00425-003-1077-5>
- GRIEU, P., RUBIN, C. & GUCKERT, A. (1995). Effect of drought on photosynthesis in *Trifolium repens*: maintenance of photosystem II efficiency and of measured photosynthesis. *Plant Physiology and Biochemistry* **33**: 19–24.
- HORTON, P. & RUBAN, A. (2005). Molecular design of the photosystem II light-harvesting antenna: photosynthesis and photoprotection. *Journal of Experimental Botany* **56**: 365–373. <http://dx.doi.org/10.1093/jxb/eri023>
- JAMES R.A., VON CAEMMERER S., CONDON A.G., MUNNS, R. & VON CAEMMERER, S. (2002). Genetic variation in tolerance to the osmotic stress component of salinity stress in durum wheat. *Functional Plant Biology* **35**: 111–123. <https://doi.org/10.1071/FP07234>

- KALAJI, H.M., GOVINDJEE, BOSA, K., KOSCIELNIAK, J. & ZUK-GOLASZEWSKA, K. (2011). Effects of salt stress on photosystem II efficiency and CO₂ assimilation of two Syrian barley landraces. *Environmental and Experimental Botany* **73**: 64–72.
<https://doi.org/10.1016/j.envexpbot.2010.10.009>
- KECK, R. & BOYER, J.S. (1974). Chloroplast response to low leaf water potentials. III. Differing inhibition of electron transport and photophosphorylation. *Plant Physiology* **53**: 474–479. <https://doi.org/10.1104/pp.53.3.474>
- LORETO, F., CENTRITTO, M. & CHARTZOULAKIS, K. (2003). Photosynthetic limitations in olive cultivars with different sensitivity to salt stress. *Plant Cell and Environment* **26**: 595–601.
<https://doi.org/10.1046/j.1365-3040.2003.00994.x>
- MEDRANO, H., ESCALONA, J.M., BOTA, J., GULÍAS, J. & FLEXAS, J. (2002). Regulation of photosynthesis of C₃ plants in response to progressive drought: stomatal conductance as a reference parameter. *Annals of Botany* **89**: 895–905.
<https://doi.org/10.1093/aob/mcf079>
- MEDRANO, H., PARRY, M.A.J., SOCÍAS, X. & LAWLOR, D.W. (1997). Long term water stress inactivates Rubisco in subterranean clover. *Annals of Applied Biology* **131**: 491–501. <https://doi.org/10.1111/j.1744-7348.1997.tb05176.x>
- MOLNÁR, I., GÁSPÁR, L., SÁRVÁRI, É., DULAI, S., HOFFMANN, B., MOLNÁR-LÁNG, M. & GALIBA, G. (2004). Physiological and morphological responses to water stress in *Aegilops biuncialis* and *Triticum aestivum* genotypes with differing tolerance to drought. *Functional Plant Biology* **31**: 1149–1159. <https://doi.org/10.1071/FP03143>
- MUNNS, R. (2002). Comparative physiology of salt and water stress. *Plant Cell and Environment* **25**: 239–250.
<https://doi.org/10.1046/j.0016-8025.2001.00808.x>
- MUNNS, R. (2005). Genes and salt tolerance: bringing them together. *New Phytologist* **167**: 645–663.
<https://doi.org/10.1111/j.1469-8137.2005.01487.x>
- MUNNS, R., JAMES, R.A. & LÄUCHLI, A. (2006). Approaches to increasing the salt tolerance of wheat and other cereals. *Journal of Experimental Botany* **57**: 1025–1043. <https://doi.org/10.1093/jxb/erj100>
- MUNNS, R. & TESTER, M. (2008). Mechanisms of salinity tolerance. *Annual Review of Plant Biology* **59**: 651–681.
<https://doi.org/10.1146/annurev.arplant.59.032607.092911>
- MURANAKA, S., SHIMIZU, K. & KATO, M. (2002). Ionic and osmotic effects of salinity on single-leaf photosynthesis in two wheat cultivars with different drought tolerance. *Photosynthetica* **40**: 201–207.
<https://doi.org/10.1023/A:1021337522431>
- NAGY, Z. & GALIBA, G. (1995). Drought and salt tolerance are not necessarily linked: a study on wheat varieties differing in drought resistance under consecutive water and salinity stresses. *Journal of Plant Physiology* **145**: 168–174.
[https://doi.org/10.1016/S0176-1617\(11\)81865-1](https://doi.org/10.1016/S0176-1617(11)81865-1)
- PRADHAN, G.P., PRASAD, P.V.V., FRITZ, A.K., KIRKHAM, M.B. & GILL, B.S. (2012). Response of *Aegilops* species to drought stress during reproductive stages of development. *Functional Plant Biology* **39**: 51–59.
<https://doi.org/10.1071/FP11171>
- QIN, J., DONG W.Y., HE K.N., YU, Y. TAN, G.D., HAN, L., DONG, M., ZHANG, Y.Y., ZHANG, D., LI, Z.A. & WANG, Z.L. (2010). NaCl salinity induced changes in water status, ion

- contents and photosynthetic properties of *Shepherdia argentea* (Pursh) Nutt. seedlings. *Plant, Soil and Environment* **56**: 325–332.
<https://doi.org/10.17221/209/2009-PSE>
- SCHNEIDER, A., MOLNÁR, I. & MOLNÁR-LÁNG, M. (2008). Utilisation of *Aegilops* (goatgrass) species to widen the genetic diversity of cultivated wheat. *Euphytica* **163**: 1–19. <https://doi.org/10.1007/s10681-007-9624-y>
- SMIRNOFF, N. (1993). The role of active oxygen in response of plants to water deficit and desiccation. *New Phytologist* **125**: 27–58.
<https://doi.org/10.1111/j.1469-8137.1993.tb03863.x>
- STILLER, I., DULAI, S., KONDRÁK, M., TARNAI, R., SZABÓ, L., TOLDI, O. & BÁNFALVI, Z. (2008). Effects of drought on water content and photosynthetic parameters in potato plants expressing the trehalose-6-phosphate synthase gene of *Saccharomyces cerevisiae*. *Planta* **227**: 299–308.
<https://doi.org/10.1007/s00425-007-0617-9>
- SUDHIR, P., POGORYELOV, D., KOVÁCS, L., GARAB, G. & MURTHY, S.D.S. (2005). The effects of salt stress on photosynthetic electron transport and thylakoid membrane proteins in the cyanobacterium *Spirulina platensis*. *Journal of Biochemistry Molecular Biology* **38**: 481–485.
<https://doi.org/10.5483/BMBRep.2005.38.4.481>
- SZOPKÓ, D., DARKÓ, É., MOLNÁR, I., KRUPPA, K., HÁLÓ, B., VOJTKÓ, A., MOLNÁR-LÁNG, M. & DULAI, S. (2017b). Photosynthetic responses of a wheat (Asakaze)-barley (Manas) 7H addition line to salt stress. *Photosynthetica* **55**: 317–328.
<https://doi.org/10.1007/s11099-016-0241-7>
- SZOPKÓ, D. & DULAI, S. (2018). Effects of drought and salt preconditioning on the heat resistance of the photosynthetic apparatus. *Acta Biologica Plantarum Agriensis* **6**: 90–107. <https://doi.org/10.21406/abpa.2018.6.90>
- SZOPKÓ, D., MOLNÁR, I., DARKÓ, É., MOLNÁR-LÁNG, M. & DULAI, S. (2017a). PEG-mediated osmotic stress responses of wheat-barley addition lines. *Acta Biologica Plantarum Agriensis* **5**(2): 3–22. <https://doi.org/10.21406/abpa.2017.5.2.3>
- TESTER, M. & DAVENPORT, R. (2003). Na⁺ tolerance and Na⁺ transport in higher plants. *Annals of Botany* **91**: 503–527. <https://doi.org/10.1093/aob/mcg058>
- TRENBERTH, K.E., SMITH, L., QIAN, T., DAI, A. & FASSULO, J. (2007). Estimates of the Global Water Budget and Its Annual Cycle Using Observational and Model Data. *Journal of Hydrometeorology* **8**: 758–769. <https://doi.org/10.1175/JHM600.1>
- VAN KOOTEN, O. & SNEL, J.F.H. (1990). The use of chlorophyll fluorescence nomenclature in plant stress physiology. *Photosynthesis Research* **25**: 147–150.
<https://doi.org/10.1007/BF00033156>
- VON CAEMMERER, S. & FARQUHAR, G.D. (1981). Some relationships between the biochemistry of photosynthesis and the gas exchange of leaves. *Planta* **153**: 376–387. <https://doi.org/10.1007/BF00384257>
- ZAHARIEVA, M., GAULIN, E., HAVAUX, M., ACEVEDO, E. & MONNEVEUX, P. (2001). Drought and heat responses in the wild wheat relative *Aegilops geniculata* Roth: Potential interest for wheat improvement. *Crop Science* **41**: 1321–1329.
<https://doi.org/10.2135/cropsci2001.4141321x>

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